



OPEN ACCESS

EDITED AND REVIEWED BY
Claudia Tarja Mierke,
Leipzig University, Germany

*CORRESPONDENCE
Rudolf E. Leube,
✉ rleube@ukaachen.de
Roy A. Quinlan,
✉ r.a.quinlan@durham.ac.uk

SPECIALTY SECTION
This article was submitted to
Cell Adhesion and Migration,
a section of the journal
Frontiers in Cell and Developmental
Biology

RECEIVED 17 January 2023
ACCEPTED 23 January 2023
PUBLISHED 13 February 2023

CITATION
Leube RE and Quinlan RA (2023), Editorial:
The wetware credentials of intermediate
filaments involves coordinating, organising
and networking in cells and tissues.
Front. Cell Dev. Biol. 11:1146618.
doi: 10.3389/fcell.2023.1146618

COPYRIGHT
© 2023 Leube and Quinlan. This is an
open-access article distributed under the
terms of the [Creative Commons
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,
distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Editorial: The wetware credentials of intermediate filaments involves coordinating, organising and networking in cells and tissues

Rudolf E. Leube^{1*} and Roy A. Quinlan^{2,3,4*}

¹Institute of Molecular and Cellular Anatomy, RWTH Aachen University, Aachen, Germany, ²Department of Biosciences, University of Durham, Upper Mountjoy Science Site, Durham, United Kingdom, ³Biophysical Sciences Institute, University of Durham, Durham, United Kingdom, ⁴Department of Biological Structure, University of Washington, Seattle, WA, United States

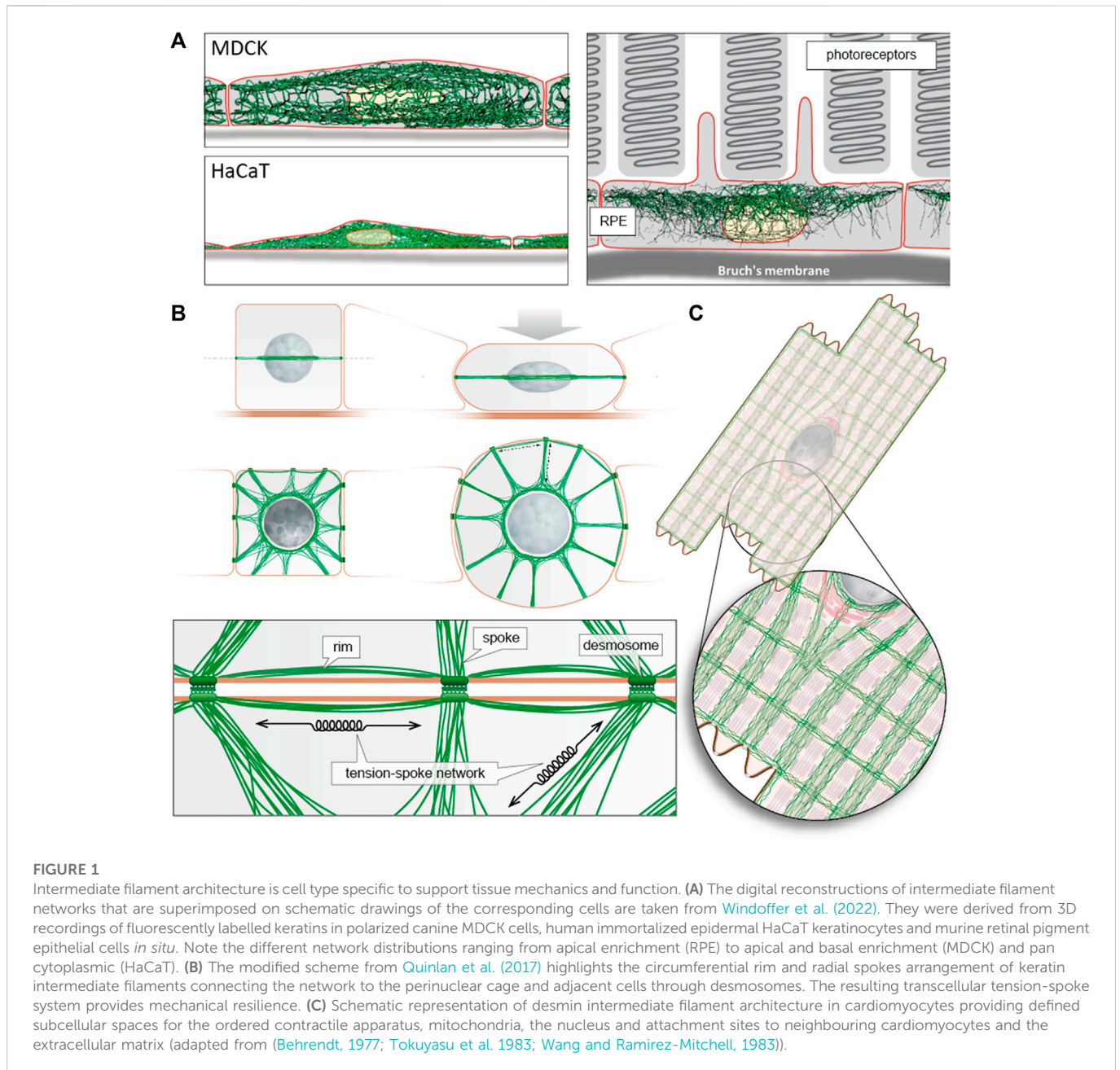
KEYWORDS

cytoskeleton, intermediate filament, cell junctions, cell shape, cell polarity and migration, wetware

Editorial on the Research Topic 3D architecture of intermediate filaments in tissue mechanics and function

In this special issue there is a collection of articles that highlight the mechano-biological signalling of and the integration of intermediate filaments within the cytoskeletal machinery. The individual and collective contribution of the individual cytoskeletal elements have been well documented (Ge et al., 2020; Serres et al., 2020; Lois-Bermejo et al., 2022; Nunes Vicente et al., 2022; Ridge et al., 2022; Sivagurunathan et al., 2022; Wu et al., 2022). Indeed the importance of the cytoskeleton as an integrated unit is accepted fully in the literature (Pegoraro et al., 2017; Hohmann and Dehghani, 2019). Intermediate filaments interconnect all subcellular compartments and they are the one cytoskeletal element where cross- β -interactions form intracellular hydrogels (Kato and McKnight, 2018) by virtue of their N- and C-terminal intrinsically disordered domains (IDDs) (Kornreich et al., 2015)—or plainly put—assist their assembly and their associated phase separation events e.g., (Li et al., 2020). It is no coincidence that previously noted “zones of exclusion” observed by conventional transmission electron microscopy (Bloise and Chacko, 1976; Borenfreund et al., 1980) should now be interpreted as evidence of their hydrogel potential e.g., nuclear pores (Fiserova et al., 2014) and cytoplasmic intermediate filament networks (Kornreich et al., 2015). The importance of these IDD to cell behaviour and to their emergent properties (Ridge et al., 2022) is a hot Research Topic in current debate. This has given rise to exciting hypotheses to explain complex cell behaviours such as motility (see the contributions by Infante and Etienne-Manneville; Kim et al., in this research topic issue) cell polarisation (ibid Despin-Guitard et al.,) epithelial-mesenchymal transitions and inflammatory responses (Ridge et al., 2022).

One such hypothesis is the “wetware” concept (Bray, 2009) as a way to conceptualise cellular and tissue decision-making at the level of individual components and processes (Kulkarni et al., 2022). Cell Biology compartmentalises systems and structures, but each function within the context of the cell and the tissue require integration and localised responses of metabolic, structural and cellular pathways. The cytoskeleton collectively provides the architecture (Figure 1) that is needed to sense, communicate and respond to the legion of stimuli received at any one time by each individual cell. It, and its associated biomolecules, can deliver the processing logic for the cell because it provides the required connections (Bray, 2009). In this respect, the intermediate filament cytoskeleton is part and



parcel of the stress response (Welch et al., 1985; Quinlan et al., 2002; Landsbury et al., 2010; Toivola et al., 2010) and to the transcriptional (Shimi and Goldman, 2014; Nazer, 2022) and to translational regulation (Magin et al., 2007; Kim and Coulombe, 2010; Mohanasundaram et al., 2022), to chaperone mediated autophagy (Bandyopadhyay et al., 2010), to respiratory efficiency (Diokmetzidou et al., 2016) and to cell division (Matsuyama et al., 2013). This identifies intermediate filaments as key interconnectors for subcellular interaction networks (Kulkarni et al., 2022). Indeed, the intermediate filament provides a surface to facilitate biomolecular folding, biomolecular complex assembly and complex organisation. Intermediate filaments as a collective provide a scale-free network across diverse length scales especially as a result of the inter-cellular organisation they afford within a tissue *via* their connection to cell-cell junctions such as the desmosome (see Green et al., in this research

topic issue). Their integrative role in mechano-signalling (Infante and Etienne-Manneville; this research topic issue) is well founded and super-resolution microscopy demonstrates that stretching filaments will reveal new, and quite possibly novel, functional nanodomains (Massou et al., 2020; Nunes Vicente et al., 2022) as also shown for lipo-oxidative stress (Lois-Bermejo et al., 2022). Stress reveals the importance of the C-terminal IDD to the biophysical properties of intermediate filaments (Aufderhorst-Roberts and Koenderink, 2019) as well as to their assembly and ultimately therefore also to cell morphology (Zhou et al., 2021).

The Research Topic highlights aspects of epithelial keratin network organization. Using a Krt8:YFP reporter mouse Desprin-Guitard and colleagues (see Despin-Guitard et al., in this research topic issue) study the keratin intermediate filament network in the developing mouse

embryo revealing a kaleidoscope of temporally and spatially determined expression profiles in embryonic and extraembryonic tissues which are interpreted as plastic adaptations of cell mechanics to growth and morphological changes. The review by Green and colleagues in this research topic issue focuses on the epidermal desmosome-keratin system as an integrator of mechanically-determined signalling. In concert with other junctions, desmosomes dictate epidermal polarization and differentiation forming a barrier by stratum-specific junctional and cytoskeletal arrangements. The authors suggest that these arrangements counteract inflammation. The paper by Yoon and colleagues in this research topic issue presents technical advancements for multidimensional and multimodal monitoring of keratin filament architecture and function. High resolution microscopy of fluorescent keratins is enabled on defined matrices and combined with traction force microscopy. In this way, the interrelationship between extracellular matrix cues with global 3D cytoskeletal network properties at the keratin filament/keratin bundle level and local forces is quantified by refined image analysis. It is further illustrated that these tools can be used for monitoring the consequences of local keratin network perturbations and ECM composition on cell mechanics in the context of transcellular network arrangement.

Infante and Etienne-Manneville in this research topic issue summarize current knowledge about the spatial arrangement and integration of cytoplasmic and nuclear intermediate filaments and their interaction with other cytoplasmic filament systems during cell migration. They emphasize the different properties of the different intermediate filament types as a basis of cell type- and function-related cellular mechanics. They further highlight the cooperativity between intermediate filaments with the other cytoskeletal systems determining motile properties of single cells and cell collectives. Direct experimental assessment of vimentin's function during metastatic invasion is finally provided by Kim and colleagues in this research topic issue. Using a novel vimentin-stabilizing drug they report on altered vimentin network morphology with consequences on adhesion and contractility resulting in cell shape changes, increased traction forces and perturbed migration.

Figure 1 presents examples of intermediate filament network organization to illustrate their function both as organizers of the subcellular space (Schwarz and Leube, 2016) and as transcellular

integrators to facilitate and support coordinated mechanical and biochemical functions in the context of tissues rather than individual cells (Hatzfeld et al., 2017). It is this framework upon which the contributions in this Research Topic have been made.

Author contributions

All authors listed have made a substantial, direct, and intellectual contribution to the work and approved it for publication.

Acknowledgments

We dedicate this editorial to Professor Werner Franke, whose passion for Intermediate Filaments and cell-cell junctions inspired many generations of cell biologists. We thank our respective academic institutions (Universities of Aachen, Durham and Washington). REL thanks the German Research Council for continued support (LE566/18-2; GRK2415/363055819). RAQ and REL thank the Biophysical Sciences Institute and the Institute of Advanced Study (Durham University) for financial support.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Aufderhorst-Roberts, A., and Koenderink, G. H. (2019). Stiffening and inelastic fluidization in vimentin intermediate filament networks. *Soft Matter* 15 (36), 7127–7136. doi:10.1039/c9sm00590k
- Bandyopadhyay, U., Sridhar, S., Kaushik, S., Kiffin, R., and Cuervo, A. M. (2010). Identification of regulators of chaperone-mediated autophagy. *Mol. Cell* 39 (4), 535–547. doi:10.1016/j.molcel.2010.08.004
- Brendt, H. (1977). Effect of anabolic steroids on rat heart muscle cells. I. Intermediate filaments. *Cell Tissue Res.* 180 (3), 303–315. doi:10.1007/BF00227598
- Blose, S. H., and Chacko, S. (1976). Rings of intermediate (100 A) filament bundles in the perinuclear region of vascular endothelial cells. Their mobilization by colcemid and mitosis. *J. Cell Biol.* 70 (21), 459–466. doi:10.1083/jcb.70.2.459
- Borenfreund, E., Schmid, E., Bendich, A., and Franke, W. W. (1980). Constitutive aggregates of intermediate-sized filaments of the vimentin and cytokeratin type in cultured hepatoma cells and their dispersal by butyrate. *Exp. Cell Res.* 127, 215–235. doi:10.1016/0014-4827(80)90428-0
- Bray, D. (2009). *Wetware: A computer in every living cell*. New Haven: Yale University Press.
- Despin-Guitard, E., Quenec'Hdu, R., Nahaboo, W., Schwarz, N., Leube, R. E., Chazaud, C., et al. (2022). Regionally specific levels and patterns of keratin 8 expression in the mouse embryo visceral endoderm emerge upon anterior-posterior axis determination. *Front. Cell Dev. Biol.* 2022 Dec 1; 10:1037041. doi:10.3389/fcell.2022.1037041
- Diokmetzidou, A., Soumaka, E., Kloukina, I., Tsikitis, M., Makridakis, M., Varela, A., et al. (2016). Desmin and α B-crystallin interplay in the maintenance of mitochondrial homeostasis and cardiomyocyte survival. *J. Cell Sci.* 129 (20), 3705–3720. doi:10.1242/jcs.192203
- Fiserova, J., Spink, M., Richards, S. A., Saunter, C., and Goldberg, M. W. (2014). Entry into the nuclear pore complex is controlled by a cytoplasmic exclusion zone containing dynamic GLFG-repeat nucleoporin domains. *J. Cell Sci.* 127 (1), 124–136. doi:10.1242/jcs.133272
- Ge, X., Zhang, T., Yu, X., Muwonge, A. N., Anandakrishnan, N., Wong, N. J., et al. (2020). LIM-nebulette reinforces podocyte structural integrity by linking actin and vimentin filaments. *J. Am. Soc. Nephrol.* 31 (10), 2372–2391. doi:10.1681/ASN.2019121261
- Green, K. J., Niessen, C. M., Rübsam, M., Perez White, B. E., and Broussard, J. A. (2022). The Desmosome-Keratin Scaffold Integrates ErbB Family and Mechanical Signaling to Polarize Epidermal Structure and Function. *Front. Cell Dev. Biol.* 10, 903696. doi:10.3389/fcell.2022.903696
- Hatzfeld, M., Keil, R., and Magin, T. M. (2017). Desmosomes and intermediate filaments: Their consequences for tissue mechanics. *Cold Spring Harb. Perspect. Biol.* 9 (6), a029157. doi:10.1101/cshperspect.a029157
- Hohmann, T., and Dehghani, F. (2019). The cytoskeleton-A complex interacting meshwork. *Cells* 8 (4), 362. doi:10.3390/cells8040362

- Kato, M., and McKnight, S. L. (2018). A solid-state conceptualization of information transfer from gene to message to protein. *Annu. Rev. Biochem.* 87, 351–390. doi:10.1146/annurev-biochem-061516-044700
- Kim, H.R., Warrington, S.J., López-Guajardo, A., Al Hennawi, K., Cook, S.L., Griffiths, Z. D. J., et al. (2022). ALD-R491 regulates vimentin filament stability and solubility, cell contractile force, cell migration speed and directionality. *Front. Cell Dev. Biol.* 10, 926283. doi:10.3389/fcell.2022.926283
- Kim, S., and Coulombe, P. A. (2010). Emerging role for the cytoskeleton as an organizer and regulator of translation. *Nat. Rev. Mol. Cell Biol.* 11 (1), 75–81. doi:10.1038/nrm2818
- Kornreich, M., Avinery, R., Malka-Gibor, E., Laser-Azogui, A., and Beck, R. (2015). Order and disorder in intermediate filament proteins. *FEBS Lett.* 589 (19), 2464–2476. doi:10.1016/j.febslet.2015.07.024
- Kulkarni, P., Bhattacharya, S., Achuthan, S., Behal, A., Jolly, M. K., Kotnala, S., et al. (2022). Intrinsically disordered proteins: Critical components of the wetware. *Chem. Rev.* 122 (6), 6614–6633. doi:10.1021/acs.chemrev.1c00848
- Landsbury, A. (2010). “Functional symbiosis between the intermediate filament cytoskeleton and small heat shock proteins,” in *Small stress proteins and human diseases*. Editors S. Simon and A. P. Arrigo, 55–87.
- Li, Y., Liu, X., Xia, C. H., FitzGerald, P. G., Li, R., Wang, J., et al. (2020). CP49 and filensin intermediate filaments are essential for formation of cold cataract. *Mol. Vis.* 26, 603–612.
- Lois-Bermejo, I., Gonzalez-Jimenez, P., Duarte, S., Pajares, M. A., and Perez-Sala, D. (2022). Vimentin tail segments are differentially exposed at distinct cellular locations and in response to stress. *Front. Cell Dev. Biol.* 10, 908263. doi:10.3389/fcell.2022.908263
- Magin, T. M., Vijayaraj, P., and Leube, R. E. (2007). Structural and regulatory functions of keratins. *Exp. Cell Res.* 313 (10), 2021–2032. doi:10.1016/j.yexcr.2007.03.005
- Massou, S., Nunes Vicente, F., Wetzel, F., Mehidi, A., Strehle, D., Leduc, C., et al. (2020). Cell stretching is amplified by active actin remodelling to deform and recruit proteins in mechanosensitive structures. *Nat. Cell Biol.* 22 (8), 1011–1023. doi:10.1038/s41556-020-0548-2
- Matsuyama, M., Tanaka, H., Inoko, A., Goto, H., Yonemura, S., Kobori, K., et al. (2013). Defect of mitotic vimentin phosphorylation causes microphthalmia and cataract via aneuploidy and senescence in lens epithelial cells. *J. Biol. Chem.* 288 (50), 35626–35635. doi:10.1074/jbc.M113.514737
- Mohanasundaram, P., Coelho-Rato, L. S., Modi, M. K., Urbanska, M., Lautenschlager, F., Cheng, F., et al. (2022). Cytoskeletal vimentin regulates cell size and autophagy through mTORC1 signaling. *PLoS Biol.* 20 (9), e3001737. doi:10.1371/journal.pbio.3001737
- Nazer, E. (2022). To be or not be (in the LAD): Emerging roles of lamin proteins in transcriptional regulation. *Biochem. Soc. Trans.* 50 (2), 1035–1044. doi:10.1042/BST20210858
- Nunes Vicente, F., Lelek, M., Tinevez, J. Y., Tran, Q. D., Pehau-Arnaudet, G., Zimmer, C., et al. (2022). Molecular organization and mechanics of single vimentin filaments revealed by super-resolution imaging. *Sci. Adv.* 8 (8), eabm2696. doi:10.1126/sciadv.abm2696
- Pegoraro, A. F., Janmey, P., and Weitz, D. A. (2017). Mechanical properties of the cytoskeleton and cells. *Cold Spring Harb. Perspect. Biol.* 9 (11), a022038. doi:10.1101/cshperspect.a022038
- Quinlan, R. A. (2002). “Cytoskeletal competence requires protein chaperones,” in *Progress in molecular and subcellular Biology. Small stress proteins*. Editors A.-P. Arrigo and W. E. G. Muller, 28, 219–233.
- Quinlan, R. A., Schwarz, N., Windoffer, R., Richardson, C., Hawkins, T., Broussard, J. A., et al. (2017). A rim-and-spoke hypothesis to explain the biomechanical roles for cytoplasmic intermediate filament networks. *J. Cell Sci.* 130 (20), 3437–3445. doi:10.1242/jcs.202168
- Ridge, K. M., Eriksson, J. E., Pekny, M., and Goldman, R. D. (2022). Roles of vimentin in health and disease. *Genes Dev.* 36 (7–8), 391–407. doi:10.1101/gad.349358.122
- Schwarz, N., and Leube, R. E. (2016). Intermediate filaments as organizers of cellular space: How they affect mitochondrial structure and function. *Cells* 5 (3), 30. doi:10.3390/cells5030030
- Serres, M. P., Samwer, M., Truong Quang, B. A., Lavoie, G., Perera, U., Gorlich, D., et al. (2020). F-actin interactome reveals vimentin as a key regulator of actin organization and cell mechanics in mitosis. *Dev. Cell* 52 (2), 210–222. doi:10.1016/j.devcel.2019.12.011
- Shimi, T., and Goldman, R. D. (2014). Nuclear lamins and oxidative stress in cell proliferation and longevity. *Adv. Exp. Med. Biol.* 773, 415–430. doi:10.1007/978-1-4899-8032-8_19
- Sivagurunathan, S., Vahabikashi, A., Yang, H., Zhang, J., Vazquez, K., Rajasundaram, D., et al. (2022). Expression of vimentin alters cell mechanics, cell-cell adhesion, and gene expression profiles suggesting the induction of a hybrid EMT in human mammary epithelial cells. *Front. Cell Dev. Biol.* 10, 929495. doi:10.3389/fcell.2022.929495
- Sun, H., Wang, A., and He, S. (2022). Temporal and spatial analysis of alzheimer’s disease based on an improved convolutional neural network and a resting-state fMRI brain functional network. *Int. J. Environ. Res. Public Health* 19 (8), 4508. doi:10.3390/ijerph19084508
- Tokuyasu, K. T., Dutton, A. H., and Singer, S. J. (1983). Immunoelectron microscopic studies of desmin (skeleton) localization and intermediate filament organization in chicken cardiac muscle. *J. Cell Biol.* 96 (6), 1736–1742. doi:10.1083/jcb.96.6.1736
- Toivola, D. M., Strnad, P., Habtezion, A., and Omary, M. B. (2010). Intermediate filaments take the heat as stress proteins. *Trends Cell Biol.* 20 (2), 79–91. doi:10.1016/j.tcb.2009.11.004
- Wang, K., and Ramirez-Mitchell, R. (1983). A network of transverse and longitudinal intermediate filaments is associated with sarcomeres of adult vertebrate skeletal muscle. *J. Cell Biol.* 96 (2), 562–570. doi:10.1083/jcb.96.2.562
- Welch, W. J., Feramisco, J. R., and Blose, S. H. (1985). The mammalian stress response and the cytoskeleton: Alterations in intermediate filaments. *Ann. N. Y. Acad. Sci.* 455 (57), 57–67. doi:10.1111/j.1749-6632.1985.tb50403.x
- Windoffer, R., Schwarz, N., Yoon, S., Piskova, T., Scholkemper, M., Stegmaier, J., et al. (2022). Quantitative mapping of keratin networks in 3D. *Elife* 11, e75894. doi:10.7554/eLife.75894
- Wu, H., Shen, Y., Sivagurunathan, S., Weber, M. S., Adam, S. A., Shin, J. H., et al. (2022). Vimentin intermediate filaments and filamentous actin form unexpected interpenetrating networks that redefine the cell cortex. *Proc. Natl. Acad. Sci. U. S. A.* 119 (10), e2115217119. doi:10.1073/pnas.2115217119
- Yoon, S., Windoffer, R., Kozyrina, A. N., Piskova, T., Di Russo, J., and Leube, R. E. (2022). Combining image restoration and traction force microscopy to study extracellular matrix-dependent keratin filament network plasticity. *plasticity. Front. Cell Dev. Biol.* 10, 901038. doi:10.3389/fcell.2022.901038
- Zhou, X., Lin, Y., Kato, M., Mori, E., Liszczak, G., Sutherland, L., et al. (2021). Transiently structured head domains control intermediate filament assembly. *Proc. Natl. Acad. Sci. U. S. A.* 118 (8), e2022121118. doi:10.1073/pnas.2022121118